

Immobilized 1,3-Dialkylimidazolium Salts as New Interface in HPLC Separation

Shu-Juan Liu, Feng Zhou, Liang Zhao, Xiao-Hua Xiao, Xia Liu, and Sheng-Xiang Jiang*
 Lanzhou Institute of Chemical Physics, Chinese Academy of Sciences,
 Tianshui Road 342, Lanzhou 730000, P. R. China

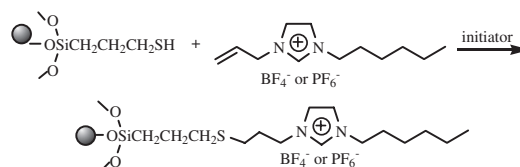
(Received December 15, 2003; CL-031235)

Silica particle chemically modified with ionic liquid is used for the first time as the stationary phase in HPLC for the separation of alkaloids following a dual mechanism.

The increasing interests in ionic liquids (ILs) largely stem from their unusual combination of properties such as environmentally benignity, nonvolatility, nonflammability.¹ So, ILs have found wide applications in various fields, e.g., as alternative solvents in routine extraction procedures,^{1,2} electrolytes in electrochemistry,³ solvents in chemical reaction,⁴ reaction media in homogeneous catalysis,⁵ and as lubricants.⁶ In the analytical science, coated IL films are employed as the stationary phase in gas chromatography (GC)⁷ and also in capillary electrophoresis (CE).⁸ Recently, they are used as the background electrolytes⁹ and especially as additives in mobile phase in HPLC.¹⁰ Surface immobilized IL has been used for heterogeneous catalysis.¹¹ However, use of surface-immobilized IL for HPLC separation has not been reported. Here we present the investigation of utilizing IL modified stationary phase as the new separation interface in HPLC.

1-Allyl-3-hexylimidazoliums ([AHIm]) combined with BF_4^- or PF_6^- counterion were prepared using the well-established method.¹² The products reacted with 3-mercaptopropyltrimethoxysilane (MPS) modified silica via the radical chain transfer addition reaction (Scheme 1) between allyl and thiol groups in the presence of azodiisobutyronitrile (AIBN).¹³ Formation of chemically immobilized layer of IL fragment on silica surface was characterized by the Raman spectrometry. The obvious Raman shift at 2578 cm^{-1} in MPS modified silica was attributed to the stretching vibration of S-H (Figure 1). After reacting with vinyl ionic liquid, its intensity decreased dramatically. The absorption bands at 3107 cm^{-1} and 3164 cm^{-1} were ascribed to the unsaturated C-H in imidazolium rings. Together with the enhanced intensity of saturated C-H between 3000 and 2800 cm^{-1} , we concluded that the IL was successfully grafted onto silica surface. The surface analysis indicated $0.58\text{ mmol}\cdot\text{g}^{-1}$ of the ILs were grafted and the coupling yield of [AHIM] salts with MPS-based silica was about 50%. IL-modified silica was packed into $150 \times 4.6\text{ mm}$ i.d. stainless steel column by slurry-packing procedure for HPLC investigation, respectively.

Alkaloids including ephedrines (1, norephedrine; 2, ephedrine and 3, pseudoephedrine, a typical pair of diastereoisomeric compounds; 4, methylephedrine) and tropane alkaloids (5, anisodamine; 6, scopolamine; 7, atropine) were chosen as the probe analytes. Ephedrines are the forbidden drugs in Olympic games and tropane alkaloids are widely used as anti-cholinergic drugs. Ephedrines are well retained on both IL-based columns with high resolution and selectivity (Figure 2a). It is worth noting that only 1% of organic solvent is added in mobile phase for an effective separation. While on C_{18} (Chromatorex C_{18} , $5\text{ }\mu\text{m}$) or C_8



Scheme 1. Preparation of ionic liquid-modified stationary phase.

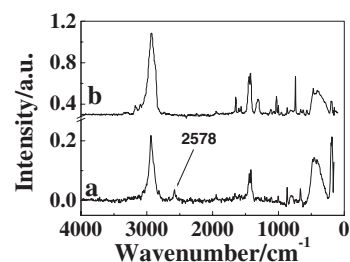


Figure 1. Raman spectra of a) MPS-modified silica and b) ionic liquid-modified silica.

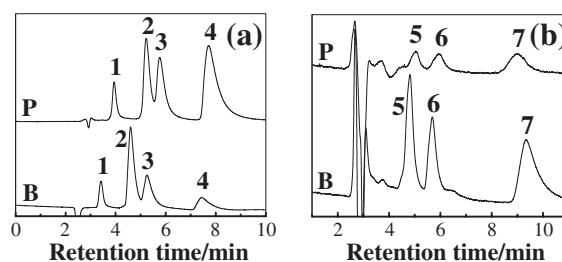


Figure 2. Typical chromatograms of (a) ephedrines and (b) tropane alkaloids on ionic liquid-modified silica. B: on [AHIm] BF_4 -based column; (a) mobile phase, 1% methanol in $0.05\text{ mol}\cdot\text{L}^{-1}\text{ KH}_2\text{PO}_4$ at pH 3.0; flowrate, $0.7\text{ mL}\cdot\text{min}^{-1}$; (b) methanol- $0.05\text{ mol}\cdot\text{L}^{-1}\text{ NaH}_2\text{PO}_4$ (10/90); flowrate, $0.6\text{ mL}\cdot\text{min}^{-1}$. P: on [AHIm] PF_6 -based column; (a) $0.02\text{ mol}\cdot\text{L}^{-1}\text{ KH}_2\text{PO}_4$; (b) methanol- $0.02\text{ mol}\cdot\text{L}^{-1}\text{ KH}_2\text{PO}_4$ (10/90); both flowrates are $0.6\text{ mL}\cdot\text{min}^{-1}$. Both the columns are $150 \times 4.6\text{ mm}$ i.d.. The data were detected at 210 nm with an UV-vis detector.

column (Zorbax Eclipse XDB- C_8 , $5\text{ }\mu\text{m}$), ephedrines exhibit very poor resolution under the similar conditions (Table 1). Comparatively, on IL-based columns the desired capacity factor (k'), superior selectivity and separation efficiency were achieved. Whereas, poor peak shapes and Rs were observed on the quaternary ammonium salts column (Table 1, SAE column). This confirms the speciality and importance of the IL-modified silica. Similarly, alkaloids 5, 6, and 7 are well separated on both IL-based columns (Figure 2b). The selectivity and the efficiency

of IL-based column have nearly no difference after repeated use, which means the good stability of it.

The pH value in mobile phase plays an important role in the separation. At lower pH, the analytes are eluted in shorter time with better peak shape, but they are retained longer at high pH. The reason is that at lower pH these basic solutes are liable to accept protons, which makes them less hydrophobic. Therefore a reversed phase mechanism may be involved in the separation. Additionally, the increase of buffer concentration could decrease the retention time by reducing the interaction between residue silanols and basic solutes.

It is seen from Figure 2 that the elution order of ephedrine on IL-based columns are the same as that on C₁₈ or C₈ column (*k'* value, showed in Table 1), indicating a similar separation mechanism. Ephedrine also have similar *k'* values on IL-based columns and on C₈ column, which is ascribed to the similar alkyl chain length (i.e., C₆ in ionic liquid, and C₈) and so they have nearly the same hydrophobicity. Though alkaloid **2** and **3** have the same chemical structures, **3** forms stable intramolecular hydrogen bond more easily than **2**, which makes it more hydrophobic and retain longer on the column than the latter. This evidence also verifies that the separation on IL-based silica follows a reversed phase mechanism that compounds with larger polarity (hydrophilicity) will be eluted sooner. The elution order of phloroglucinol < hydroquinone < phenol or trimesic acid < *p*-phthalic acid < benzoic acid on the IL-based columns also confirms a reversed phase mechanism. It is concluded that long substituted alkyl chains and the hydrophobic counter ions make ionic liquid-modified silica the reversed phase stationary phases.

Alkaloid **5** is eluted before **6** on IL modified columns while on C₁₈ or C₈ column, it displays an inverse order (see *k'* value, Table 1). It is well known that alkaloid **6** has higher hydrophilic property than **5**, which leads to a smaller retention time of **6** on C₁₈ or C₈ column. However, when adding sodium dodecyl sulfate (SDS) as the ion pair agent to the mobile phase, **5** and **6** are eluted in the same order on C₁₈ column as that on IL-based column.¹⁴ Though alkaloid **6** has less basicity than **5** because of the steric hindrance effect, the oxonium salt formed at lower pH on the epoxy group of alkaloid **6** could form ion pair with the pairing ion, resulting in its stronger retention on C₁₈ column. The same situation can be expected on IL-based column that at lower pH the formation of ion pair between oxonium salt of alkaloid **6** and the BF₄⁻ or PF₆⁻ anion confined on silica leads to a stronger retention of **6**. So ion-pairing interaction plays an important role in the separation on these new packings. In addition, it has been proved that ionic liquids possess ion pair characteristic and can form ion-pairing complex with analytes in electroanalysis.¹⁵

In a word, this kind of IL-modified stationary phases display characteristics not only of reversed phase chromatography but also ion-pair chromatography. The analytes can be effectively separated on IL-based stationary phase without addition of ion pair agent¹⁶ or triethylamine¹⁷ in mobile phase as that on C₁₈ column.

In summary, chemically immobilized IL on silica surface is utilized as a new interface for the HPLC separation of alkaloids. Effective separation is ascribed to both the hydrophobicity and the ionic property of IL-modified stationary phases. Considering the large family of ionic liquids with tunable substituted alkyl chains and counter anions, IL-based stationary phase has great

Table 1. Comparison of *k'* (capacity factor) and Rs (resolution) on varied columns

| Comp. | [AHIm]BF ₄ | | [AHIm]PF ₆ | | C ₁₈ | | C ₈ | | SAE | |
|----------|------------------------|-----|------------------------|-----|------------------------|-----|------------------------|-----|------------------------|-----|
| | ^a <i>k'</i> | Rs | ^b <i>k'</i> | Rs | ^c <i>k'</i> | Rs | ^d <i>k'</i> | Rs | ^e <i>k'</i> | Rs |
| 1 | 0.46 | | 0.48 | | 6.5 | | 0.46 | | 0.46 | |
| 2 | 1.0 | 3.2 | 0.97 | 3.8 | 13 | 5.1 | 0.95 | 3.6 | 0.51 | 0.4 |
| 3 | 1.2 | 1.4 | 1.2 | 1.3 | 17 | 2.6 | 1.3 | 2.8 | 0.61 | 0.5 |
| 4 | 2.2 | 2.9 | 1.9 | 2.6 | 18 | 0.9 | 1.3 | 0.6 | 0.81 | 0.9 |
| | a2 | | b2 | | c2 | | d2 | | e2 | |
| 5 | 0.86 | | 0.92 | | 4.0 | | 0.76 | | 0.54 | |
| 6 | 1.2 | 1.6 | 1.3 | 1.3 | 2.9 | 2.4 | 0.45 | 2.8 | 0.55 | 0.2 |
| 7 | 2.6 | 3.8 | 2.4 | 3.0 | 7.6 | 4.6 | 1.7 | 5.2 | 0.72 | 2.0 |

a1: mobile phase, 1% methanol in 0.02 mol·L⁻¹ KH₂PO₄; a2, b2, e2: methanol-0.02 mol·L⁻¹ NaH₂PO₄ (10/90); b1: 0.03 mol·L⁻¹ KH₂PO₄; c1: water at pH 3.0 adjusted with H₃PO₄; c2: 20% methanol in 0.02 mol·L⁻¹ KH₂PO₄; d1: 0.02 mol·L⁻¹ KH₂PO₄; d2: 10% methanol in 0.02 mol·L⁻¹ KH₂PO₄; e1: 0.01 mol·L⁻¹ NaH₂PO₄.

variety and can be investigated further in HPLC. Moreover, the letter presents a new addition to the versatile applications of IL.

References

- J. G. Huddleston, H. D. Willauer, R. P. Swatoski, A. E. Visser, and R. D. Rogers, *Chem. Commun.*, **1998**, 1765.
- J. F. Liu, G. B. Jiang, Y. G. Chi, Y. Q. Cai, Q. X. Zhou, and J. T. Hu, *Anal. Chem.*, **75**, 5870 (2003).
- H. Ohno, C. Suzuki, K. Fukumoto, M. Yoshizawa, and K. Fujita, *Chem. Lett.*, **32**, 450 (2003).
- J. S. Yadav, B. V. S. Reddy, A. K. Basak, and A. V. Narsaiah, *Chem. Lett.*, **32**, 988 (2003); J. S. Yadav, B. V. S. Reddy, and M. Srinivas, *Chem. Lett.*, **32**, 1060 (2003).
- T. Itoh, E. Akasaki, and Y. Nishimura, *Chem. Lett.*, **2002**, 154; T. Itoh, N. Ouchi, S. Hayase, and Y. Nishimura, *Chem. Lett.*, **32**, 654 (2003).
- C. F. Ye, W. M. Liu, Y. X. Chen, and L. G. Yu, *Chem. Commun.*, **2001**, 2244.
- D. W. Armstrong, L. F. He, and Y. S. Liu, *Anal. Chem.*, **71**, 3873 (1999); J. L. Anderson and D. W. Armstrong, *Anal. Chem.*, **75**, 4851 (2003).
- W. D. Qin and S. F. Y. Li, *Analyst*, **128**, 37 (2003).
- E. G. Yanes, S. R. Gratz, M. J. Baldwin, S. E. Robison, and A. M. Stalcup, *Anal. Chem.*, **73**, 3838 (2001); M. Vaher, M. Koel, and M. Kaljurand, *J. Chromatogr., A*, **979**, 27 (2002); W. D. Qin, H. P. Wei, and S. F. Y. Li, *J. Chromatogr., A*, **985**, 447 (2003).
- L. J. He, W. Z. Zhang, L. Zhao, X. Liu, and S. X. Jiang, *J. Chromatogr., A*, **1007**, 39 (2003).
- C. P. Mehnert, R. A. Cook, N. C. Dispenziere, and M. Afeworki, *J. Am. Chem. Soc.*, **124**, 12932 (2002).
- M. Hirao, K. Ito, and H. Ohno, *Electrochim. Acta*, **45**, 1291 (2000).
- C. E. Song, J. S. Lim, S. C. Kim, K. J. Lee, D. Y. Chi, *Chem. Commun.*, **2000**, 2415; F. Zhou, W. M. Liu, M. Chen, and D. C. Sun, *Chem. Commun.*, **14**, 2446 (2001).
- L. Y. He, G. D. Zhang, and Y. Y. Tong, *J. Chromatogr.*, **481**, 428 (1989).
- A. J. Fry, *J. Electroanal. Chem.*, **546**, 35 (2003).
- K. Sagara, T. Oshima, and T. Misaki, *Chem. Pharm. Bull.*, **31**, 2359 (1983).
- C. Imaz, D. Carreras, R. Navajas, C. Rodriguez, A. F. Rodriguez, J. Maynar, and R. Cortes, *J. Chromatogr., A*, **631**, 201 (1993).